

Amendments to the Specification

Please replace the paragraph appearing at page 5, lines 16-22 with the following amended paragraph:

D1
Figure 3 shows the alignment of HrpW (SEQ ID NO: 2) with pectate lyases of *Nectria haematococca*, mating type VI (*Fusarium solani* f. sp. *pisi*) (PIA-Nh = SEQ ID NO: 4; PIB-Nh = SEQ ID NO: 5; PIC-Nh = SEQ ID NO: 6; PID-Nh = SEQ ID NO: 7) and of *Erwinia carotovora* subsp. *carotovora* (Pel-3-Ec = SEQ ID NO: 8; PelB-Ec = SEQ ID NO: 9). The sequences were aligned by the PILEUP program (GCG software package, Version 7.3) with default parameters, and an alignment was manually edited using LINEUP program in the same package. Conserved residues are boxed, highly conserved regions are underlined, and potential α -helices in HrpW are shaded. A consensus (SEQ ID NO: 10) within the Pel domain is shown below the alignment.

Please replace the paragraph appearing at page 24, lines 16-28 with the following amended paragraph (note that underlined titles were underlined in the original):

D2
E. amylovora Ea321Rp (a rifampicin-resistant derivative of Ea321), Ea321-K49 (*hrpL*::Tn 10-miniKm) (Wei, et al., J. Bacteriol., 177:6201-10 (1995), which is hereby incorporated by reference), Ea321-G84 (*hrcC*::Tn5-*gusA1*) (Kim et al., J. Bacteriol., 179:1690-97 (1997), which is hereby incorporated by reference), Ea273Rp, Ea273-K49, and Ea273-G73 (*hrcV*::Tn5-*gusA1*) were grown overnight in Terrific broth, transferred to a *hrp* minimal medium (Huynh, et al., Science, 345:1374-77 (1989), which is hereby incorporated by reference) at ~~1×10^8~~ 1×10^8 cfu/ml, and incubated at 20°C until the bacteria grew to ~~1×10^9~~ 1×10^9 cfu/ml. Cultures were centrifuged at 17,500 g and the pellet was resuspended in a loading buffer. The supernatant was passed through a membrane filter (0.2 μ m pore size; Whatman Inc., Fairfield, NJ) after adding 1mM PMSF, and concentrated 100-fold using Centricon-10 and Microcon-10 (Amicon, Inc., Beverly, MA) at 4°C. Both the cell and supernatant fractions were then subjected to SDS-PAGE in a 10% gel.

Please replace the paragraph appearing at page 25, lines 13-22 with the following amended paragraph (note that underlined titles were underlined in the original):

D3
Elicitation of the HR was tested by infiltrating protein or bacterial preparations into the intercellular space of leaves of tobacco (*Nicotiana tabacum* L. 'xanthi') and other

D3 conc
plants (Kim, et al., J. Bacteriol., 179:1690-97 (1997), which is hereby incorporated by reference). Cells were grown either in Luria broth (*E. coli* DH5 α and MC4100) or a *hrp* minimal medium (*E. amylovora* Ea321 and Ea321-T5) (Huynh, et al., Science, 345:1374-77 (1989), which is hereby incorporated by reference) to ~~5H10⁸~~ 5 x 10⁸ cfu/ml, and resuspended in 5 mM KPO₄ buffer (pH 6.5) to ~~2H10⁸~~ 2 x 10⁸ cfu/ml (*E. coli* strains) or ~~5H10⁸~~ 5 x 10⁸ cfu/ml (*E. amylovora* strains). Inhibitors of plant metabolism used included cycloheximide at 100 μ M, LaCl₃ at 1 mM, and Na₃VO₄ at 50 μ M.

Please replace the paragraph appearing at page 25, lines 25-31 with the following amended paragraph:

D4
Genomic DNA was digested with *Eco*RI, electrophoresed on a 0.7% agarose gel, transferred to an Immobilon-N membrane (Millipore Co., Bedford, MA), and hybridized with the ³²P-Labelled 1.4kb-*Hpa*I fragment of pCPP1227 at 65°C for 24 hr in a hybridization solution of 6X SSC, 5X Denhardt's reagent, 0.5% SDS, and 100 μ g/ml denatured fragmented salmon sperm DNA (Sambrook et al., Molecular Cloning: A Laboratory Manual, (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY) (1989) at 9.31-9.57. The membrane was washed twice with a solution of ~~2H SSC~~ 2X SSC and 1.0% SDS at 65°C, and washed with ~~0.1H SSC~~ 0.1X SSC until no radioactivity is detected in the wash solution. For low stringency hybridizations, the membrane was incubated at 50°C and washed with ~~2H SSC~~ 2X SSC at 45°C.
